





# Exploring Viral Metagenomics in ASFV-Positive Samples from the Philippines Reveals Co-Infection With Other Swine Pathogens

<u>Ann Catherine Y. Cabrera</u><sup>1</sup>, Allan Michael C. Mamites<sup>1</sup>, Gabriela Ilona B. Janairo<sup>1</sup>, Jake C. Mauhay<sup>1</sup>, Ma Jowina H. Galarion<sup>1</sup>, Brian Schwem<sup>1</sup>, Coleen M. Pangilinan<sup>1</sup>, Jennifer L. Maravilla<sup>2</sup>, Brent Kristian D. Molina<sup>2</sup>, Rachel R. Azul<sup>2</sup>, and Rohani C. Navarro<sup>1,3</sup>

National Institute of Molecular Biology and Biotechnology, National Institutes of Health, University of the Philippines Manila, Manila, PHILIPPINES
 Animal Disease Diagnosis and Reference Laboratory, Bureau of Animal Industry, Department of Agriculture, Quezon City, PHILIPPINES
 National Training Center for Biosafety and Biosecurity, National Institutes of Health, University of the Philippines Manila, Manila, PHILIPPINES

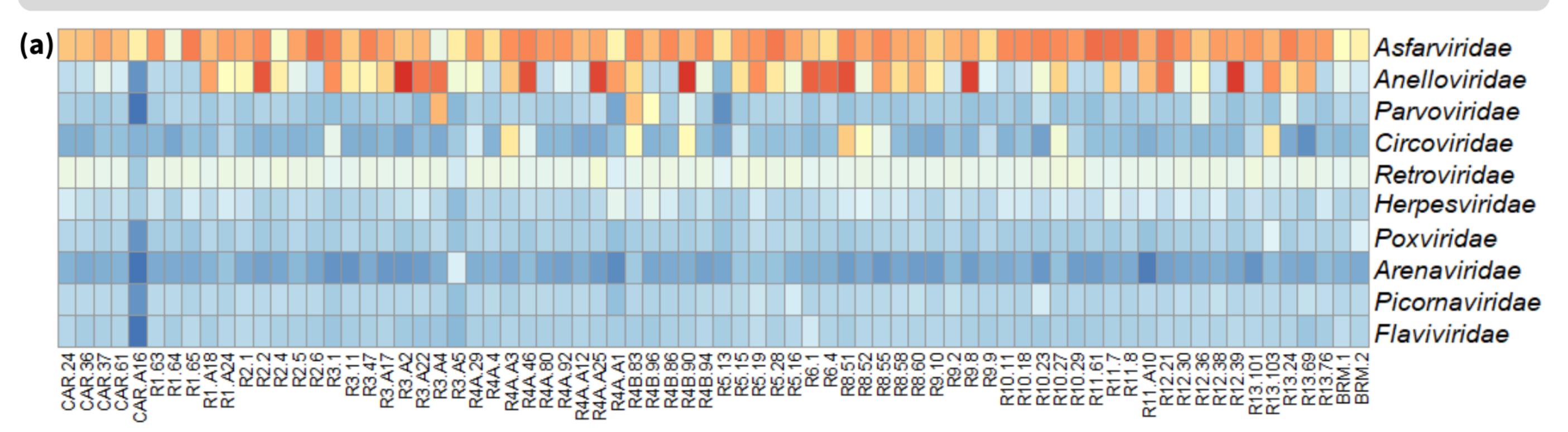
## INTRODUCTION

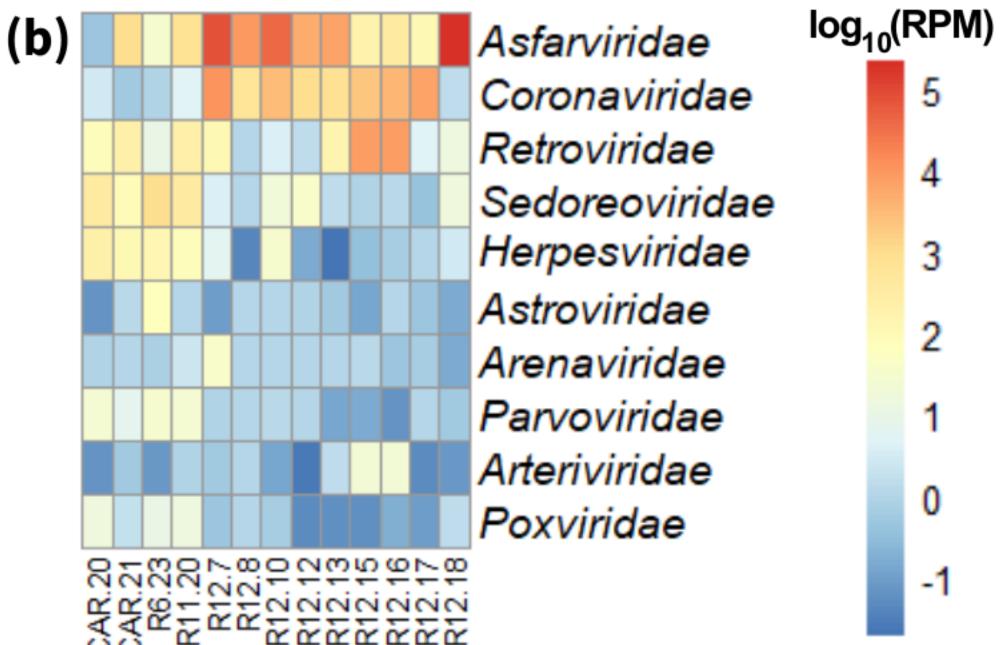
Co-infections are common in swine and can influence disease outcomes. While co-infections of various swine pathogens are increasingly recognized, the presence of other DNA or RNA viruses in ASF-infected swine remains largely uncharacterized.

**OBJECTIVE**: To identify co-infecting swine viruses in ASF-positive samples using a metagenomic sequencing approach

# ASF-positive sample collection DNA / RNA extraction and pre-processing Illumina Sequencing from 15 Philippine regions QC, Assembly, and Annotation Taxonomic classification

# RESULTS AND DISCUSSION





**Figure 1**. Heatmaps displaying the relative abundance of the top 10 vertebrate-associated viral families (y-axis) identified through k-mer-based taxonomic classification using Kraken2 on (a) DNA sequencing and (b) RNA sequencing data, presented on a  $\log_{10}$  reads per million (RPM) scale across samples. The color intensity corresponds to the  $\log_{10}(\text{RPM})$  values, as shown in the legend. The x-axis refers to sample IDs representing samples collected from individual pigs from different regions of the Philippines.

**Table 1.** List of viruses with near to complete-length genome assemblies from DNA and RNA sequencing.

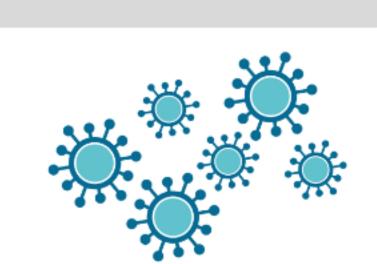
Method	Family	Genus	Species	Contigs	Length (kb)	Prevalence	Seen with ASFV before?
DNA-seq /	Anelloviridae	Iotatorquevirus	lotatorquevirus suida1a	159	1.5 - 5.8	58.67%	Yes <sup>1</sup>
		Kappatorquevirus	Kappatorquevirus suidak2a	29	1.5 - 2.8	20%	No
			Kappatorquevirus suidak2b	16	1.5 - 3.0	21.33%	No
	Circoviridae	Circovirus	Circovirus porcine2	17	1.7	21.33%	Yes <sup>2,3</sup>
	Parvoviridae	Tetraparvovirus	Tetraparvovirus ungulate2	1	5.2	1.33%	Yes <sup>4</sup>
		Copiparvovirus	Copiparvovirus ungulate2	1	5.6	1.33%	No
			Copiparvovirus ungulate4	3	3.4 - 5.7	4%	No
RNA-seq Sedoreoviridae Rotavirus			Rotavirus alphagastroenteritidis	6*	0.5 - 2.3	14.29%	No

\*Near to complete-length sequences of this virus' coding regions were assembled

In summary, 63 out of 75 (84%) DNA samples and 2 out of 14 (14.29%) RNA samples showed the presence of at least one viral species other than ASFV.

### CONCLUSION

To our knowledge, this is the first report to explore co-infection in ASFV-infected swine using an unbiased sequencing approach.





These findings provide baseline data on swine viral diversity to aid in further investigations of viral co-infections in correlation with clinical signs.

# ACKNOWLEDGEMENTS









